

Dr Jan-Ulrich Kreft

Lecturer in Computational Biology

School of Biosciences (</schools/biosciences/index.aspx>)

Contact details

Telephone [+44 \(0\)121 41 48851](tel:+441214148851) ([tel:+44 121 41 48851](tel:+441214148851))

Fax +44 (0)121 41 48844

Email j.kreft@bham.ac.uk (<mailto:j.kreft@bham.ac.uk>)

Centre for Systems Biology, School of Biosciences
University of Birmingham

Edgbaston
Birmingham
B15 2TT
UK



About

Dr Jan-Ulrich Kreft is a microbiologist turned mathematical modeller.

Lab website address: www.biosciences-labs.bham.ac.uk/kreftlab/index.html (<http://www.biosciences-labs.bham.ac.uk/kreftlab/index.html>)

For an up-to-date publication list see my [Google Scholar page](http://scholar.google.co.uk/citations?user=hLRsYpsAAAAJ&hl=en) (<http://scholar.google.co.uk/citations?user=hLRsYpsAAAAJ&hl=en>)

Feedback and office hours

My office is in the Centre for Systems Biology, click here for [directions to the CSB \(findUs.html\)](#)

Qualifications

- 1991 Diplom Biologe, University of Tübingen, Germany
- 1995 Dr rer nat (PhD), University of Konstanz, Germany

Biography

- 10/1985 – 08/1991 Studied Biology at the Universities of Konstanz and Tübingen, Germany
- 09/1991 – 06/1995 PhD in Microbiology at the University of Konstanz, Germany
- 06/1995 – 02/1997 Postdoc with Professor Bernhard Schink at the University of Konstanz, Germany
- 03/1997 – 08/1998 DFG fellowship with Professor Julian Wimpenny at the University of Cardiff, UK
- 09/1998 – 08/2001 Postdoctoral Research Associate (BBSRC) with Professor Julian Wimpenny at the University of Cardiff, UK
- 04/1999 – 06/1999 Guest scientist in the lab of Professor Mark van Loosdrecht, TU Delft, NL
- 09/2001 – 08/2007 Wissenschaftlicher Assistent in the Theoretical Biology group of Professor Wolfgang Alt, University of Bonn, Germany
- 09/2007 – present Lecturer in Computational Biology, University of Birmingham, UK

Teaching

Undergraduate Teaching

- Bio107 (Enzymes and Metabolism): I introduce Microbiology
- Bio153 (Microbiology and Infectious Disease): I teach Microbial Lifestyles
- Bio234 (Ecology): I teach Microbial Symbioses with Plants and Animals
- Bio262 (Membranes, Energy and Metabolism): I teach Microbial Bioenergetics
- Bio303 (Applied and Environmental Microbiology): I teach Microbial Ecology

Postgraduate Teaching

- Statistics for PhD students
- Essentials of Biology for Mathematicians
- Masterclass in Agent-based Modelling for PhD students
- Individual-based Modelling of Microbial Interactions & Processes Using iDyNoMiCS – one week course for PhD students and Postdocs

Postgraduate supervision

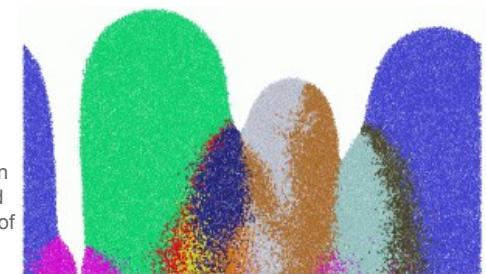
Research

Research Theme within School of Biosciences: Molecular Microbiology

Lab website address: www.biosciences-labs.bham.ac.uk/kreftlab/index.html (<http://www.biosciences-labs.bham.ac.uk/kreftlab/index.html>)

Cooperation and communication of microbes in biofilms, metabolic division of labour, individual-based modelling, systems biology Dynamics of interaction

I'm broadly interested in the dynamics of interaction between 'parts' (e.g. individual organisms) and how these interactions give rise to emergent behaviour on the next higher level of organisation (e.g. the population). There are far more fascinating examples than one can possibly study, so I have focused on competition, cooperation, and communication of microbes in spatially structured systems such as biofilms.



How the simple process of substrate diffusion and biomass growth leads to complex biofilm structures can be seen in the image of a nitrifying biofilm above, which shows different clones of ammonia oxidizers in different colours and all clones of nitrite oxidizers in magenta. The competition for oxygen diffusing in from above leads to the formation of finger-like structures and lateral inhibition (in this case suffocation). This biofilm structure was generated by an Individual-based Model that describes the behaviour and activities of the individual microbes and how they interact with each other and their environment. This model treats each individual bacterial cell as a hard ball, following the spirit of considering a spherical cow. See the [individual-based modelling of biofilms paper](http://mic.sgmjournals.org/cgi/content/abstract/147/11/2897) (<http://mic.sgmjournals.org/cgi/content/abstract/147/11/2897>).

Below are some examples of my current research projects, for more details and other projects please visit [the Kreft lab pages](http://www.biosciences-labs.bham.ac.uk/kreftlab/index.html) (<http://www.biosciences-labs.bham.ac.uk/kreftlab/index.html>).

Evolution of cooperation in biofilms

We are studying a case of cooperation that consists in the economic use of resources, known as the 'tragedy of the commons' and have shown that this cooperation can evolve in biofilms. The benefit of saving common and limiting resources is indiscriminately but locally shared among the neighbourhood while the more efficient metabolism comes at the cost of a reduced specific growth rate of the cooperating individuals due to a thermodynamic trade-off between biomass yield per amount of resource and growth rate.

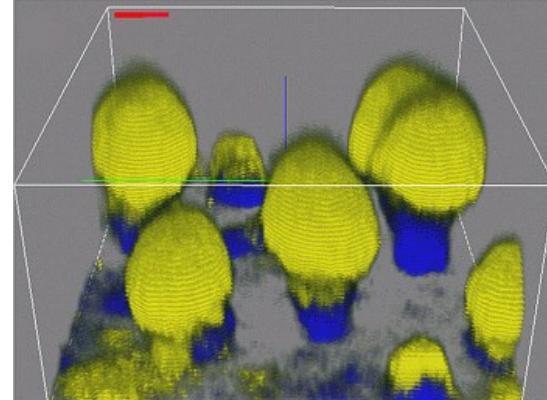


Using an individual-based model, the competition between wasteful (blue) and efficient (red) individual bacteria can be simulated in spatially complex systems such as biofilms.

It turns out that spatial structure is of key importance for the success of the slower but more efficient bacteria. The importance of spatial structure can be best appreciated watching [movies of model simulations](http://www.theobio.uni-bonn.de/people/jan_kreft/altruism.html) (http://www.theobio.uni-bonn.de/people/jan_kreft/altruism.html), for further explanations please see the [altruism paper](http://www.theobio.uni-bonn.de/downloads/jan_kreft/papers/kreft2004.pdf) (http://www.theobio.uni-bonn.de/downloads/jan_kreft/papers/kreft2004.pdf).

Twitching motility and biofilm structure

Some bacteria like *Pseudomonas aeruginosa* can pull themselves across surfaces by the use of type IV pili as grappling hooks ([more on Alex Merz's web site](http://www.webcom.com/alexey/moviepage.html) (<http://www.webcom.com/alexey/moviepage.html>)). This twitching motility called process has remarkable effects on biofilm structure in *P. aeruginosa* because a wild type population phenotypically separates into an immotile and motile subpopulation.



The immotile subpopulation forms clonal microcolony stalks and the motile subpopulation appears to crawl up on the stalks to form caps, allowing the population as a whole to form the mushroom structures you can see in the micrograph (the image is from a biofilm including wild type in yellow and an immotile mutant in blue for a more reliable separation of the subpopulations).

This phenotypic split has some interesting evolutionary consequences we have briefly discussed in a recent [review](http://dx.doi.org/10.1111/j.1574-6968.2006.00280.x) (<http://dx.doi.org/10.1111/j.1574-6968.2006.00280.x>).

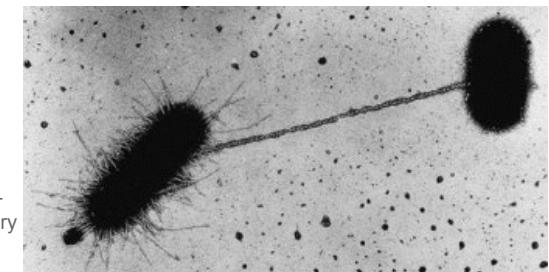
We have been developing an individual-based model of biofilms including twitching motility to dissect the processes involved and to understand the fitness effects of this behaviour, for a description see our [paper](http://dx.doi.org/10.2166/wst.2007.275) (<http://dx.doi.org/10.2166/wst.2007.275>).

This project has been a collaboration with [Cristian Picioroanu](http://www.biofilms.bt.tudelft.nl/) (<http://www.biofilms.bt.tudelft.nl/>), at the Delft University of Technology, The Netherlands, and [Mikkel Klausen, Janus Haagensen, Tim Tolker-Nielsen, Søren Molin](http://www.cbm.biocentrum.dtu.dk/) (<http://www.cbm.biocentrum.dtu.dk/>), at the Danish Technical University in Lyngby near Copenhagen.

Horizontal gene transfer (HGT) in biofilms

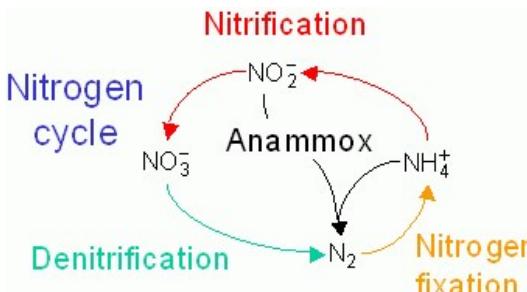
The horizontal transfer of genes from individual to individual by conjugation (see electron microscopic image by Charles C. Brinton, Jr., of a mating pair initially brought together by means of an F pilus), or via extracellular DNA by transformation, is a remarkable and prevalent phenomenon in bacterial communities, where the spatial arrangement of donor and recipient is obviously important.

This calls for the use of spatially explicit models that allow the status and activities of the individuals to change due to gene transfer, in other words, individual-based models. We are developing such an individual-based model of conjugation that we will use to predict HGT in biofilms and also to investigate the evolutionary and ecological consequences of HGT.



This project is a collaboration with Barth F Smets' group at the Danish Technical University in Lyngby near Copenhagen. Our 'next-generation' individual-base model we have been developing for this and many future projects is called [individual-based simulator for Dynamics of Microbial Communities Simulator \(iDyMoMiCS\)](http://emerg.er.dtu.dk/index.php?option=com_content&task=view&id=17&Itemid=18) (http://emerg.er.dtu.dk/index.php?option=com_content&task=view&id=17&Itemid=18). It was originally developed by Laurent Lardon and then Brian Merkey, with contributions to the design and/or programming by Sonia Martins, Cristian Picioroanu, Andreas Dötsch, Joao Xavier, Barth F Smets, and myself.

One-step nitrification



Since Winogradsky's discovery in 1890, it is known that nitrification, an important link in the global Nitrogen cycle, is carried out by two physiologically and phylogenetically distinct groups of bacteria, the first step is the incomplete oxidation of ammonia (NH_4^+) to nitrite (NO_2^-), which is oxidized further to nitrate (NO_3^-) by a second group of bacteria. Stahl and coworkers recently discovered that the first step can also be carried out by archaea.

Why doesn't the first group of bacteria or archaea oxidize ammonia completely? We have argued that this metabolic division of labour can be explained by kinetic theory of optimal pathway length, which assumes that the production of enzymes as well as the presence of intermediates is costly and that the number of ATP generating steps is proportional to the length of the pathway. From these assumptions, the existence of an optimal pathway length follows which maximises the rate of ATP production. Shortening the pathway by oxidizing ammonia only incompletely could therefore increase the rate of ATP production and growth, increasing fitness in environments such as enrichment cultures that select for fastest growth rate.

However, there is a trade-off between growth rate and growth yield, since the longer pathway would have more ATP generating steps, thereby increasing growth yield, which we have shown (see the cooperation section above) to be advantageous when bacteria grow in clonal clusters as is typical for biofilms. We therefore postulate the existence of bacteria that completely oxidize ammonia to nitrate and have suggested to use biofilm cultures to select for the higher yield that these "lithotrophs missing in nature" (Broda, 1977) should have. We are interested in isolating and constructing such bacteria by genetic engineering and *in silico*.

See our [opinion paper for an explanation of kinetic theory and further examples of microbial food webs with versus without division of labour](#). (<http://dx.doi.org/10.1016/j.tim.2006.03.006>) A further paper has analyzed the conditions under which a food-chain carrying out complete ammonia oxidation is advantageous compared to incomplete ammonia oxidation (see partial nitrification paper).

This project has been a collaboration with Julio Pérez, Dpt. d'Enginyeria Química, [Universitat Autònoma de Barcelona, Spain](#) (<http://www.uab.es/>).

Cell-cell communication

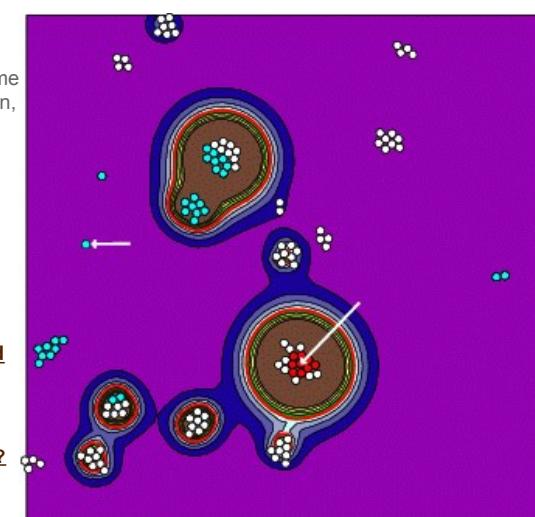
Bacteria are simple organisms without brains, but they nevertheless evolved some fascinating social behaviours, for example cell-cell communication by production of diffusible signal molecules called autoinducers. This has been studied a lot in well-mixed liquid cultures, where the concentration of autoinducers depends on the density of cells, and this has led to the idea that cells indeed use these autoinducers for the control of gene expression by the density of cells in a population, known as quorum sensing. However, in most natural habitats, the complex physical structure of the environment and the clustered distribution of cells growing on surfaces (think soil), can have a stronger effect on the concentration of autoinducers a given cell experiences than the density of cells.

The figure shows results of a simulation, using iDynoMiCS, by my former student Andreas Dötsch. The distribution of cells (circles) is clustered and the contours show the autoinducer concentration (colour scheme as in a topographic map: blue is below threshold (below sea level), red contour line = threshold concentration, green and brown = islands of upregulation, the shore is green and mountains are brown). You can see that small clusters of autoinducer producing cells easily become upregulated while lone cells don't. Further, the autoinducer that the focal red cell in the microcolony experiences has been produced by itself and its close neighbours only (all the red cells), while the focal lone cell in cyan listens mostly to itself and a variety of other cells, all in cyan.

This shows that communication can be kept private to microcolonies, and that has important evolutionary consequences because microcolonies are clones, all cells are offspring of a single founding cell.

See our [opinion paper for a critical analysis of quorum sensing problems in the natural habitat and our proposed solution](#) (<http://dx.doi.org/10.1038/nrmicro1600>).

This project has been a collaboration with my colleagues **Burkhard A. Hense**, (<http://ibb.gsf.de/ag3/person.php?name=Burkhard+Hense>) **Christina Kuttler** (<http://ibb.gsf.de/ag3/person.php?name=Christina+Kuttler>), and **Johannes Müller** (<http://ibb.gsf.de/ag3/person.php?name=Johannes+M%C3%83%C2%BCller>) from the **Institute for Biomathematics and Biometry (IBB)** (<http://ibb.gsf.de/>) and **Michael Rothbauer** (http://www.gsf.de/amp/rhizo/Seiten/ThMR_eng.html) and **Anton Hartmann** (http://www.gsf.de/amp/rhizo/Seiten/ThAH_eng.html) from the **Department Microbe-Plant-Interaction** (http://www.gsf.de/amp/rhizo/Seiten/Main_eng.htm), both based at the **GSF - National Research Center for Environment and Health** (http://www.gsf.de/index_en.php), near München, Germany.



Other activities

- Library Representative for Biosciences
- Biosciences Graduate School Committee Member
- Deputy Director, Centre for Systems Biology
- Member, Faculty of 1000, Environmental Microbiology Section
- Editor, FEMS Microbiology Letters

Publications

Please see my [Google Scholar page](#) (<http://scholar.google.co.uk/citations?user=hLRsYpsAAAAJ&hl=en>) for further or more recent publications

If you don't have access to one of these publications, just email me for a pdf (j.kreft@bham.ac.uk)

- Kreft JU, Schink B (1993). [Demethylation and degradation of phenylmethylethers by the sulfide-methylating homoacetogenic bacterium strain TMBS4](#). (<http://dx.doi.org/10.1007/BF00290912>) Archives of Microbiology 159: 308-315
- Kreft JU, Schink B (1994). [O-Demethylation by the homoacetogenic anaerobe Holophaga foetida studied by a new photometric methylation assay using electrochemically produced cob\(I\)alamin](#). (<http://dx.doi.org/10.1111/j.1432-1033.1994.00945.x>) European Journal of Biochemistry 226: 945-951
- Liesack W, Bak F, Kreft JU, Stackebrandt E (1994). [Holophaga foetida gen. nov., sp. nov., a new, homoacetogenic bacterium degrading methoxylated aromatic compounds](#). (<http://dx.doi.org/10.1007/BF00264378>) Archives of Microbiology 162: 85-90
- Kappler O, Janssen PH, Kreft JU, Schink B (1997). [Effects of alternative methyl group acceptors on the growth energetics of the O-demethylating](#)

- anaerobe *Holophaga foetida*. (<http://dx.doi.org/10.1099/00221287-143-4-1105>) *Microbiology* 143: 1105-1114
- Kreft JU, Schink B (1997). [Specificity of O-demethylation in extracts of the homoacetogenic *Holophaga foetida* and demethylation kinetics measured by a coupled photometric assay.](#) (<http://dx.doi.org/10.1007/s002030050456>) *Archives of Microbiology* 167: 363-368
- Kreft JU, Booth G, Wimpenny JWT (1998). [BacSim, a simulator for individual-based modelling of bacterial colony growth.](#) (<http://dx.doi.org/10.1099/00221287-144-12-3275>) *Microbiology* 144: 3275-3287
- Kreft JU, Picioreanu C, Wimpenny JWT, van Loosdrecht MCM (2001). [Individual-based modelling of biofilms.](#) (<http://mic.sgmjournals.org/cgi/content/abstract/147/11/2897>) *Microbiology* 147: 2897-2912
- Kreft JU, Wimpenny JWT (2001). [Effect of EPS on biofilm structure and function as revealed by an individual-based model of biofilm growth.](#) (<http://www.iwaponline.com/wst/04306/wst043060135.htm>) *Water Science and Technology* 43(6): 135-141
- van Loosdrecht MCM, Heijnen JJ, Eberl HJ, Kreft JU, Picioreanu C (2002). [Mathematical modelling of biofilm structures.](#) (<http://dx.doi.org/10.1023/A:1020527020464>) *Antonie van Leeuwenhoek* 81: 245-256
- Picioreanu C, Kreft JU, van Loosdrecht MCM (2004). [Particle-based multidimensional multispecies biofilm model.](#) (<http://dx.doi.org/10.1128/AEM.70.5.3024-3040.2004>) *Applied and Environmental Microbiology* 70: 3024-3040
- Kreft JU (2004). [Biofilms promote altruism.](#) (<http://dx.doi.org/10.1099/mic.0.26829-0>) *Microbiology* 150: 2751-2760
- Kreft JU (2004). [Conflicts of interest in biofilms.](#) (<http://dx.doi.org/10.1017/S1479050504001486>) *Biofilms* 1: 265-276
- Kreft JU, Bonhoeffer S (2005). [The evolution of groups of cooperating bacteria and the growth rate versus yield trade-off.](#) (<http://dx.doi.org/10.1099/mic.0.27415-0>) *Microbiology* 151: 637-641
- Grijspolder K, Kreft JU, Messens W (2005). [Individual-based modelling of growth and migration of *Salmonella enteritidis* in hen's eggs.](#) (<http://dx.doi.org/10.1016/j.ijfoodmicro.2004.10.028>) *International Journal of Food Microbiology* 100: 323-333
- Dens EJ, Bernaerts K, Standaert AR, Kreft JU, Van Impe JF (2005). [Cell division theory and individual-based modeling of microbial lag. Part II.](#) (<http://dx.doi.org/10.1016/j.ijfoodmicro.2004.11.017>) Modeling lag phenomena induced by temperature shifts. *International Journal of Food Microbiology* 101: 319-332
- Costa E, Pérez J, Kreft JU (2006). [Why is metabolic labour divided in nitrification?](#) (<http://dx.doi.org/10.1016/j.tim.2006.03.006>) *Trends in Microbiology* 14: 213-219
- Klausen M, Gjermansen M, Kreft JU, Tolker-Nielsen T (2006). [Dynamics of development and dispersal in sessile microbial communities: examples from *Pseudomonas aeruginosa* and *Pseudomonas putida* model biofilms.](#) (<http://dx.doi.org/10.1111/j.1574-6968.2006.00280.x>) *FEMS Microbiology Letters* 261: 1-11
- Hense BA, Kuttler C, Müller J, Rothbauer M, Hartmann A, Kreft JU (2007). [Does efficiency sensing unify quorum and diffusion sensing?](#) (<http://dx.doi.org/10.1038/nrmicro1600>) *Nature Reviews Microbiology* 5: 230-239
- Picioreanu C, Kreft JU, Klausen M, Haagensen JA, Tolker-Nielsen T, Molin S (2007). [Microbial motility involvement in biofilm structure formation – a 3D modelling study.](#) (<http://dx.doi.org/10.2166/wst.2007.275>) *Water Science and Technology* 55: 337-343
- Dötsch A, Severin J, Alt W, Galinski EA, Kreft JU (2008). [A mathematical model for growth and osmoregulation in halophilic bacteria.](#) (<http://dx.doi.org/10.1099/mic.0.2007/012237-0>) *Microbiology* 154: 2956-2969
- Schuster S, Kreft JU, Schroeter A, Pfeiffer T (2008). [Use of game-theoretical methods in biochemistry and biophysics.](#) (<http://dx.doi.org/10.1007/s10867-008-9101-4>) *Journal of Biological Physics* 34: 1-17
- Pérez J, Costa E, Kreft JU (2009). [Conditions for partial nitrification in biofilm reactors and a kinetic explanation.](#) (<http://dx.doi.org/10.1002/bit.22249>) *Biotechnology and Bioengineering* 103: 282-295
- Kreft JU (2009). [Mathematical modeling of microbial ecology: spatial dynamics of interactions in biofilms and guts.](#) (<http://estore.asm.org/viewItemDetails.asp?itemID=851>) In *Foodborne Microbes: Shaping the Host Ecosystem*. Jaykus LA, Wang HH, Schlesinger LS (eds), pp 347-377. Washington, DC: ASM Press
- Bock M, Tyagi AK, Kreft JU, Alt W (2010). [Generalized Voronoi tessellation as a model of two-dimensional cell tissue dynamics.](#) (<http://dx.doi.org/10.1007/s11538-009-9498-3>) *Bulletin of Mathematical Biology* 72: 1696-1731
- Schuster S, Kreft JU, Brenner N, Wessely F, Theißßen G, Ruppel E, Schroeter A (2010). [Cooperation and cheating in microbial exoenzyme production - Theoretical analysis for biotechnological applications.](#) (<http://dx.doi.org/10.1002/biot.200900303>) *Biotechnology Journal* 5: 751-758
- Lardon LA, Merkey BV, Martins S, Dötsch A, Picioreanu C, Kreft JU, Smets BF (2011). [iDyNoMiCS: next-generation individual-based modelling of biofilms.](#) (<http://dx.doi.org/10.1111/j.1462-2920.2011.02414.x>) *Environmental Microbiology* 13: 2416-2434
- Costello C, Kreft JU, Thomas CM, Mendes PM (2011). Protein nanoarrays for high resolution patterning of bacteria on gold surfaces. In: Toms S, Weil R (eds) *Nanoproteomics: Methods and Protocols*. Springer Protocols, Methods in Molecular Biology Series. Humana Press, New York
- Merkey BV, Lardon LA, Seoane JM, Kreft JU, Smets BF (2011). [Growth dependence of conjugation explains limited plasmid invasion in biofilms: an individual-based modelling study.](#) (<http://dx.doi.org/10.1111/j.1462-2920.2011.02535.x>) *Environmental Microbiology* 13: 2435-2452
- Schmidt SI, Picioreanu C, Craenen B, Mackay R, Kreft JU, Theodoropoulos G (2011). [A multi-scale agent-based distributed simulation framework for groundwater pollution management.](#) (<http://dx.doi.org/10.1109/DS-RT.2011.33>) Proceedings of the 15th IEEE International Symposium on Distributed Simulation and Real Time Applications (DS-RT'11), Salford, Manchester, UK, September 4-7 2011. (<http://c4i.gmu.edu/events/conferences/2011/DS-RT/>) IEEE Computer Society, Los Alamitos, CA, USA, pp 18-27
- Costello CM, Kreft JU, Thomas CM, Hammes DM, Bao P, Evans SD, Mendes PM (2012). [Exploiting additive and subtractive patterning for spatially controlled and robust bacterial co-cultures.](#) (<http://dx.doi.org/10.1039/C2SM26111A>) *Soft Matter* 8: 9147-9155
- van der Wal A, Tecon R, Kreft JU, Mooij WM, Leveau JHJ (2013). [Explaining bacterial dispersion on leaf surfaces with an individual-based model \(PHYLLOSIM\).](#) (<http://dx.doi.org/10.1371/journal.pone.0075633>) *PLoS ONE* 8: e75633
- Kreft JU, Plugge CM, Grimm V, Prats C, Leveau JHJ, Banitz T, Baines S, Clark J, Ros A, Klapper I, Topping CJ, Field AJ, Schuler A, Litchman E, Hellweger FL (2013). [Mighty small: Observing and modeling individual microbes becomes big science.](#) (<http://dx.doi.org/10.1073/pnas.1317472110>) *PNAS* 110: 18027-18028